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POSTER ABSTRACTS

508.BONE MARROW FAILURE: ACQUIRED

Distinct Immune and Molecular Profiles of Hepatitis-Associated Aplastic Anemia

Roma V. Rajput, MD¹, Dalton Hironaka, BA¹, Ruba Shalhoub², Lemlem Alemu¹, Emma M. Groarke, MD³, Jennifer Lotter¹, Olga Rios¹, Ivana Darden, RN¹, Colin O. Wu, PhD², Fernanda Gutierrez-Rodrigues, PhD⁴, Xingmin Feng, PhD¹, Neal S. Young, MD¹, Bhavisha A. Patel, MD¹

¹Hematology Branch, National Heart, Lung, and Blood Institute, National Institutes of Health, Bethesda, MD

²Office of Biostatistics Research, National Heart, Lung, and Blood Institute, National Institutes of Health, Bethesda, MD

³Hematology Branch, National Heart, Lung, and Blood Institute, NIH, Bethesda, MD

⁴Hematology Branch, National Heart, Lung, and Blood Institute, National Institute of Health, Bethesda, MD

Background: Hepatitis-associated aplastic anemia (HAAA) is characterized by the development of hematopoietic failure within months following an acute episode of non-infectious hepatitis. Although disease presentation with pancytopenia and a hypocellular marrow in HAAA is similar to non-hepatitis immune AA (non-HAAA), the immune and molecular profile in HAAA has not been completely elucidated. Here, we aimed to assess HAAA patients for cytokines and presence of clonal events linked to AA, the presence of paroxysmal nocturnal hemoglobinuria (PNH) clones and somatic mutations in peripheral blood mononuclear cells (PBMC) subpopulations, and compared to a non-HAAA cohort.

Methods: A cohort of HAAA patients (n=30), age and sex-matched non-HAAA patients (n=24) and healthy controls (n=23) seen at the National Institutes of Health since 2003 were retrospectively screened for a panel of inflammatory cytokines and growth factors by Luminex, and somatic mutations in sorted CD3-, CD3+CD4+, and CD3+CD8+ PBMC fractions by error-corrected DNA sequencing. A customized panel with 42 myeloid-related and 49 autoimmunity-related genes at minimum limited of detection of 0.1% were used. All samples were before any immunosuppressive treatment (IST). Cytokine levels and frequencies of PNH clones and clonal hematopoiesis (CH) in different fractions were compared among groups.

Results: HAAA patients were in average young (median age=23); 37% were at age < 18 (Table 1). Although disease severity and blood counts were similar to the non-HAAA cohort, immune and molecular markers of immune AA were distinct among cohorts. The frequency of PNH clones was significantly lower in HAAA compared to the non-HAAA cohort (15% v 58%, p=0.0008). The median size of PNH clones in HAAA and non-HAAA was 1.4% and 7.4%, respectively (p =0.03). The cytokine profiles performed in the sera of 22 HAAA before IST were marked by significantly higher levels of IL-1ra (p=0.0008), CXCL-10 (p=0.0038), and VEGF (p=0.0023) in comparison to 31 non-HAAA assessed patients; IL-6 (p=0.0002) and CD40L (p=0.0001) levels were significantly lower than in non-HAAA. As reported in immune AA, HAAA patients had higher levels of TPO (p=0.0001), G-CSF (p=0.0001), and EPO (p=0.0001) than controls; lower levels of CCL-5 (p=0.0001) and CCL-11 (p=0.0025) were seen in the HAAA. FLT3-L levels were markedly elevated in both AA cohorts when compared to HC.

The clonal landscape was also strikingly different between HAAA and non-HAAA in all fractions evaluated (Figure 1). In HAAA, 3 out of 21 assessed patients (14%) had CH in the CD3- fraction at maximum variant allele frequency (VAF) of 1.8%; one patient had two *RUNX1* mutations while a *JAK2* and *DNMT3A* mutation were found in the other two patients. No mutations were found in CD4+ and CD8+ compartments in subjects with HAAA. In contrast, 13 of 24 non-HAAA patients (54%) had CH in the CD3- populations, which was dominated by *PIGA, BCOR*, and *DNMT3A* clones, a pattern consistent with the known clonal landscape of immune AA. The frequency of non-HAAA with CH in the CD4+ and CD8+ compartments was 7/24 (29%) and 8/24 (30%) patients, respectively; CH in these fractions was similar to those found in CD3- dominated by mutations in *PIGA, BCOR*, and *DNMT3A* at VAFs ranging from 0.2% to 15% (Figure 1). CH in lymphoid-related genes was only found in a single patient with a *STAT3* p.S614Rmutation at VAF of 3% exclusive to the CD8+ fraction.

Conclusion: HAAA has a distinct immunologic and molecular profile than non-HAAA patients. PNH clones were less frequent in HAAA and patterns of cytokines were significantly different than the non-HAAA cohort. CH in *PIGA* and *BCOR*, highly associated with AA, were not present in our small HAAA cohort. Our results indicate a distinct underlying pathophysiology that initiates the immune destruction resulting in marrow failure in HAAA patients. Confirming these findings in a larger cohort and correlating with clinical outcomes is ongoing.

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Disclosures No relevant conflicts of interest to declare.

Table 1: Demographics and Clinical Characteristics of Hepatitis-associated aplastic anemia (HAAA) and non-hepatitis associated aplastic anemia (non-HAAA) cohorts.					Figure 1	: Clonal landscape of	HAAA versus non-h	epatitis AA. Oncoprin	t of 45 patients with av	ailable ECS. Abbreviatio	ons: HAAA, hepatitis-
Number of patients (n,	HAAA(N=30)	Non-HAAA (N=24)	Healthy Control (N=23)	P value			associated	aplastic anemia; ECS,	error-corrected sequer	ncing.	
Gender				0.7							
Male	20 (67%)	16 (67%)	13 (57%)				Hepatitis AA (HAAA)			on-Hepatitis AA (non-HA/	
Female	10 (33%)	8 (33%)	10 (43%)								
Age, years (median, range)			N 12	0.92**		CD3 negative	CD3+ CD4+	CD3+ CD8+	CD3 negative	CD3+CD4+	CD3+CD8+
<18	11 (37%)	10 (42%)	0			CODECCE PARAMAX STAR		BOCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCCC	00000000000000000000000000000000000000	C0388088C-01/20808000	10000660000-00/11 x # C-00
18-39	16 (53%)	11 (46%)	13 (57%)		1	3333333333333333333333333333	******************	45333333333333333333333333333	**********************	**********************	********************
>40	3 (10%)	3 (13%)	10 (43%)			*******************	*********************	*******************	332333333333332333333333333333	333555555555555555555555555555555555555	*****
Race				0.39	Photo close > 1%-5%	CONTRACTOR OF A DESCRIPTION OF A DESCRIP					
White	20 (67%)	12 (50%)	11(48%)								
Black or African American	5 (17%)	8 (33%)	8 (35%)		PNH clone >5%						
Asian	1.(3%)	1 (4%)	3 (13%)			the second state of the second state	THE REPORT OF THE REPORT OF THE	the second second second second second			and the second se
Mattiple Race/Unknown	4 (13%)	3 (13%)	1(4%)		PIGA		111229357878782531112832				CONTRACTOR CONTRACTOR
Severity of Disease											
SAA	13 (43%)	13 (54%)			- DNMT3A	• • • • • • • • • •					
VSAA	16 (53%)	11(46%)			TET2						
MAA	1 (3%)	0			BCORAL		100000000000000000000000000000000000000				
GPI negative neutrophil by flow (> 1%)	3/27 (15%)	14/24 (58%)		0.0008	ASXL						
GPI negative neutrophil clone size (VAF, %)	1,4%	7.4%		0.03	RUNX1						
Pre-treatment blood counts (median, range)					JAK2						
ANC (K/uL)	0.19 (0.0-1.2)	0.26 (0.0-0.7)		0.82	KRAS						
PLT (K)(L)	10.5 (1-47)	8 (1-22)		0.11	PHES						
ARC (KAL)	14.6 (2.4-104)	17.3 (1.6-60.4)		0.47					• • • • • • • • • • • • • • • • • • •	•	
Somatic Mutations*					STAG2		110102002000000000000000000000000000000				
CD3- (# of mutations)	4	13	3	0.005	TP53						
CD4+ (# of mutations)	0	7	2	0.01	UZAFI				• •	•	
CD8+ (# of mutations)	0	8	4	0.01			110000000000000000000000000000000000000				
severe aplastic anemia; VS/	VA, very severe aplar cy: ANC, absolute i	stic anemia; MAA, moderale	of HAAA and non-HAAA or explastic anemia: GPL gives elet count; ARC, absolute	sylphosphatidylinositol	ri un				I	1	•



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